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(54) Title: HUMAN THETA SUBUNIT OF THE GABA-A RECEPTOR

(57) Abstract

The present invention relates to the cloning of a novel cDNA sequence encoding the theta receptor subunit of the GABAA receptor; to stably co-transfected eukaryotic cell lines capable of expressing a GABAA receptor, which receptor comprises the novel theta receptor subunit; and to the use of such cell lines in screening for and designing medicaments which act upon the GABAA receptor.

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WO 98/49293 - 1 - PCT/GB98/01206

HUMAN THETA SUBUNIT OF THE GABA-A RECEPTOR

This invention concerns the cloning of a novel cDNA sequence encoding a particular subunit of the human GABAA receptor. In addition, the invention relates to a stable cell line capable of expressing said cDNA and to the use of the cell line in a screening technique for the design and development of subtype-specific medicaments.

Gamma-amino butyric acid (GABA) is a major inhibitory neurotransmitter in the central nervous system. It mediates fast synaptic inhibition by opening the chloride channel intrinsic to the GABAA receptor. This receptor comprises a multimeric protein of molecular size 230-270 kDa with specific binding sites for a variety of drugs including benzodiazepines, barbiturates and β-carbolines, in addition to sites for the agonist ligand GABA (for reviews see MacDonald and Olsen, *Ann. Rev. Neurosci.*, 1994, 17, 569; and Whiting *et al*, *Int. Rev. Neurobiol.*, 1995, 38, 95).

Molecular biological studies demonstrate that the receptor is composed of several distinct types of subunit, which are divided into four classes (α , β , γ and δ) based on their sequence similarities. To date, in mammals, six types of α (Schofield et al., Nature (London), 1987, 328, 221; Levitan et al., Nature (London), 1988, 335, 76; Ymer et al., EMBO J., 1989, 8, 1665; Pritchett & Seeberg, J. Neurochem., 1990, 54, 802; Luddens et al., Nature (London), 1990, 346, 648; and Khrestchatisky et al., Neuron, 1989, 3, 745), three types of β (Ymer et al., EMBO J., 1989, 8, 1665), three types of γ (Ymer et al., EMBO J., 1990, 9, 3261; Shivers et al., Neuron, 1989, 3, 327; and Knoflach et al, FEBS Lett., 1991, 293, 191) and one δ subunit (Shivers et al., Neuron, 1989, 3, 327) have been identified. More recently, a further member of the GABA receptor gene family, ϵ , has been identified (Davies et al., Nature, 1997, 385, 820). The polypeptide is 506 amino acids in length and exhibits greatest amino acid sequence identity

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with the GABA_A receptor γ_3 subunit (47%), although this degree of homology is not sufficient for it to be classified as a fourth γ subunit.

The differential distribution of many of the subunits has been characterised by in situ hybridisation (Shivers et al., Neuron, 1989, 3, 327; Wisden et al, J. Neurosci., 1992, 12, 1040; and Laurie et al, J. Neurosci, 1992, 12, 1063) and this has permitted it to be speculated which subunits, by their co-localisation, could theoretically exist in the same receptor complex.

Various combinations of subunits have been co-transfected into cells to identify synthetic combinations of subunits whose pharmacology 10 parallels that of bona fide GABAA receptors in vivo (Pritchett et al., Science, 1989, 245, 1389; Pritchett and Seeberg, J. Neurochem., 1990, 54, 1802; Luddens et al., Nature (London), 1990, 346, 648; Hadingham et al, Mol. Pharmacol., 1993, 43, 970; and Hadingham et al., Mol. Pharmacol., 1993, 44, 1211). This approach has revealed that, in addition to an α and 15 β subunit, either γ_1 or γ_2 (Pritchett et al., Nature (London), 1989, 338, 582; Ymer et al., EMBO J., 1990, 9, 3261; and Wafford et al., Mol. Pharmacol., 1993, 44, 437) or γ₃ (Herb et al., Proc. Natl. Acad. Sci. USA, 1992, 89, 1433; Knoflach et al., FEBS Lett., 1991, 293, 191; and Wilson-Shaw et al., FEBS Lett., 1991, 284, 211) is also generally required to confer 20 benzodiazepine sensitivity, and that the benzodiazepine pharmacology of the expressed receptor is largely dependent on the identity of the α and γ subunits present. Receptors containing a δ subunit (i.e. $\alpha\beta\delta$) do not appear to bind benzodiazepines (Shivers et al., Neuron, 1989, 3, 327; and 25 Quirk et al., J. Biol. Chem., 1994, 269, 16020). Combinations of subunits have been identified which exhibit the pharmacological profile of a BZ1 type receptor $(\alpha_1\beta_1\gamma_2)$ and a BZ₂ type receptor $(\alpha_2\beta_1\gamma_2 \text{ or } \alpha_3\beta_1\gamma_2, \text{ Pritchett } \text{et}$ al., Nature (London), 1989, 338, 582), as well as GABAA receptors with a novel pharmacology, α₅β₂γ₂ (Pritchett and Seeberg, J. Neurochem., 1990, 30 54, 1802), $\alpha_4\beta_2\gamma_2$ (Wisden et al, FEBS Lett., 1991, 289, 227) and $\alpha_6\beta_2\gamma_2$

WO 98/49293 - 3 - PCT/GB98/01206

(Luddens et al., Nature (London), 1990, 346, 648). The pharmacology of these expressed receptors appears similar to that of those identified in brain tissue by radioligand binding, and biochemical experiments have begun to determine the subunit composition of native GABA receptors (McKernan & Whiting, Tr. Neurosci., 1996, 19, 139). The exact structure of receptors in vivo has yet to be definitively elucidated.

The present invention relates to a new class of GABA receptor subunit, hereinafter referred to as the theta subunit (θ subunit).

The nucleotide sequence for the theta subunit, together with its deduced amino acid sequence corresponding thereto, is depicted in Figure 1 of the accompanying drawings.

The present invention accordingly provides, in a first aspect, a DNA molecule encoding the theta subunit of the human GABA receptor comprising all or a portion of the sequence depicted in Figure 1, or a modified human sequence.

In an alternative aspect, the present invention provides a DNA molecule encoding the theta subunit of the human GABA receptor comprising all or a portion of the sequence depicted in Figure 2, or a modified human sequence.

The term "modified human sequence" as used herein referes to a variant of the DNA sequences depicted in Figure 1 and Figure 2. Such variants may be naturally occurring allelic variants or non-naturally occurring or "engineered" variants. Allelic variation is well known in the art in which the nucleotide sequence may have a substitution, deletion or addition of one or more nucleotides without substantial alteration of the function of the encoded polypeptide. Particularly preferred allelic variants arise from nucleotide substitution based on the degeneracy of the genetic code.

The sequencing of the novel cDNA molecules in accordance with the invention can conveniently be carried out by the standard procedure described in accompanying Example 1; or may be accomplished by

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alternative molecular cloning techniques which are well known in the art, such as those described by Maniatis et al. in Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Press, New York, 2nd edition, 1989.

In a further aspect, the present invention also relates to polynucleotides (for example, cDNA, genomic DNA or synthetic DNA) which hybridize under stringent conditions to the DNA molecules depicted in Figure 1 and Figure 2. As used herein, the term "stringent conditions" will be understood to require at least 95% and preferably at least 97% identity between the hybridized sequences. Polynucleotides which hybridize under stringent conditions to the DNA molecules depicted in Figure 1 and Figure 2 preferably encode polypeptides which exhibit substantially the same biological activity or function as the polypeptides depicted in Figure 1 and Figure 2, respectively.

The present invention further relates to a GABA theta subunit polypeptide which has the deduced amino acid sequence of Figure 1 or Figure 2, as well as fragments, analogs and derivatives thereof.

The terms "fragment", "derivative" and "analog" when referring to the polypeptide of Figure 1 or Figure 2, means a polypeptide which retains essentially the same biological activity or function as the polypeptide depicted in Figure 1 or Figure 2. Thus, an analog may be, for example, a proprotein which can be activated by cleavage of the proprotein portion to produce an active mature polypeptide.

The polypeptide of the present invention may be a recombinant polypeptide, a natural polypeptide or a synthetic polypeptide, preferably a recombinant polypeptide.

The fragment, derivative or analog of the polypeptide of Figure 1 or Figure 2 may be one in which one or more of the amino acid residues are substituted with a conserved or non-conserved amino acid residue (preferably a conserved amino acid residue) and such substituted amino acid residues may or may not be one encoded by the genetic code; or one in

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WO 98/49293 - 5 - PCT/GB98/01206

which one or more of the amino acid residues includes a substituent group; or one in which the mature polypeptide is fused with another compound, such as a compound to increase the half-life of the polypeptide (for example, polyethylene glycol); or one in which the additional amino acids are fused to the mature polypeptide, such as a leader or secretory sequence or a sequence which is employed for purification of the mature polypeptide or a proprotein sequence. Such fragments, derivatives and analogs are deemed to be within the technical capabilities of those skilled in the art.

The polypeptides and DNA molecules of the present invention are preferably provided in an isolated form, and preferably are purified to homogeneity.

The term "isolated" means that the material is removed from its original environment (e.g., the natural environment if it is naturally occurring). For example, a naturally-occurring DNA molecule or polypeptide present in a living animal is not isolated, but the same DNA molecule or polypeptide, separated from some or all of the coexisting materials in the natural system, is isolated. Such DNA molecules could be part of a vector and/or such DNA molecules or polypeptides could be part of a composition, and still be isolated in that such vector or composition is not part of its natural environment.

In another aspect, the invention provides a recombinant expression vector comprising the nucleotide sequence of the human GABA receptor theta subunit together with additional sequences capable of directing the synthesis of the said human GABA receptor theta subunit in cultures of stably co-transfected eukaryotic cells.

The term "expression vectors" as used herein refers to DNA sequences that are required for the transcription of cloned copies of recombinant DNA sequences or genes and the translation of their mRNAs in an appropriate host. Such vectors can be used to express eukaryotic genes in a variety of hosts such as bacteria, blue-green algae, yeast cells,

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insect cells, plant cells and animal cells. Specifically designed vectors allow the shuttling of DNA between bacteria-yeast, bacteria-plant or bacteria-animal cells. An appropriately constructed expression vector should contain: an origin of replication for autonomous replication in host cells, selective markers, a limited number of useful restriction enzyme sites, a high copy number, and strong promoters. A promoter is defined as a DNA sequence that directs RNA polymerase to bind to DNA and to initiate RNA synthesis. A strong promoter is one which causes mRNAs to be initiated at high frequency. Expression vectors may include, but are not limited to, cloning vectors, modified cloning vectors, specifically designed plasmids or viruses.

The term "cloning vector" as used herein refers to a DNA molecule, usually a small plasmid or bacteriophage DNA capable of self-replication in a host organism, and used to introduce a fragment of foreign DNA into a host cell. The foreign DNA combined with the vector DNA constitutes a recombinant DNA molecule which is derived from recombinant technology. Cloning vectors may include plasmids, bacteriophages, viruses and cosmids.

The recombinant expression vector in accordance with the invention may be prepared by inserting the nucleotide sequence of the GABA theta subunit into a suitable precursor expression vector (hereinafter referred to as the "precursor vector") using conventional recombinant DNA methodology known from the art. The precursor vector may be obtained commercially, or constructed by standard techniques from known expression vectors. The precursor vector suitably contains a selection marker, typically an antibiotic resistance gene, such as the neomycin or ampicillin resistance gene. The precursor vector preferably contains a neomycin resistance gene, adjacent the SV40 early splicing and polyadenylation region; an ampicillin resistance gene; and an origin of replication, e.g. pBR322 ori. The vector also preferably contains an inducible promoter, such as MMTV-LTR (inducible with dexamethasone)

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WO 98/49293 - 7 - PCT/GB98/01206

or metallothionin (inducible with zinc), so that transcription can be controlled in the cell line of this invention. This reduces or avoids any problem of toxicity in the cells because of the chloride channel intrinsic to the GABAA receptor.

One suitable precursor vector is pMAMneo, available from Clontech Laboratories Inc. (Lee et al., Nature, 1981, 294, 228; and Sardet et al., Cell, 1989, 56, 271). Alternatively the precursor vector pMSGneo can be constructed from the vectors pMSG and pSV2neo.

The recombinant expression vector of the present invention is then produced by cloning the GABA receptor theta subunit cDNA into the above precursor vector. The receptor subunit cDNA is subcloned from the vector in which it is harboured, and ligated into a restriction enzyme site, e.g. the Hind III site, in the polylinker of the precursor vector, for example pMAMneo or pMSGneo, by standard cloning methodology known from the art, and in particular by techniques analogous to those described herein. Before this subcloning, it is often advantageous, in order to improve expression, to modify the end of the theta subunit cDNA with additional 5' untranslated sequences, for example by modifying the 5' end of the theta subunit DNA by addition of 5' untranslated region sequences from the α_1 subunit DNA. Alternatively, expression of the theta subunit cDNA may be modified by the insertion of an epitope tag sequence such as c-myc.

According to a further aspect of the present invention, there is provided a stably co-transfected eukaryotic cell line capable of expressing a GABA receptor, which receptor comprises the theta receptor subunit, at least one alpha receptor subunit and optionally one or more beta, gamma, delta, or epsilon receptor subunit.

This is achieved by co-transfecting cells with multiple expression vectors, each harbouring cDNAs encoding for an α , θ , and optionally one or more β , γ , δ , or ϵ GABA receptor subunits. In a further aspect, therefore, the present invention provides a process for the preparation of a eukaryotic cell line capable of expressing a GABA receptor, which

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comprises stably co-transfecting a eukaryotic host cell with at least two expression vectors, one such vector harbouring the cDNA sequence encoding the theta GABA receptor subunit, and another such vector harbouring the cDNA sequence encoding an alpha GABA receptor subunit. The stable cell-line which is established expresses an $\alpha\theta$ GABA receptor.

Each receptor thereby expressed, comprising a unique combination of α , θ and optionally one or more subunits selected from β , γ , δ or ϵ subunits, will be referred to hereinafter as a GABA receptor "subunit combination".

Expression of the GABA receptor may be accomplished by a variety of different promoter-expression systems in a variety of different host cells. The eukaryotic host cells suitably include yeast, insect and mammalian cells. Preferably the eukaryotic cells which can provide the host for the expression of the receptor are mammalian cells. Suitable host cells include rodent fibroblast lines, for example mouse Ltk-, Chinese hamster ovary (CHO) and baby hamster kidney (BHK); HeLa; and HEK293 cells. It is necessary to incorporate at least one α subunit, the θ subunit, and optionally one or more subunits selected from β , γ , δ or ε into the cell line in order to produce the required receptor. Within this limitation, the choice of receptor subunit combination is made according to the type of activity or selectivity which is being screened for.

In order to employ this invention most effectively for screening purposes, it is preferable to build up a library of cell lines, each with a different combination of subunits. Typically a library of 5 or 6 cell line types is convenient for this purpose. Preferred subunit combinations include: $\alpha\theta\beta$, $\alpha\theta\gamma$, $\alpha\theta\delta$, and $\alpha\theta\epsilon$, and most especially $\alpha_1\theta\gamma_2$. Further preferred subunit combinations include $\alpha\beta\theta\gamma$ and $\alpha\beta\theta\epsilon$, and most especially $\alpha_2\beta_1\theta\gamma_1$ and $\alpha_2\beta_3\theta\gamma_2$.

Cells are then co-transfected with the desired combination of the expression vectors. There are several commonly used techniques for transfection of eukaryotic cells *in vitro*. Calcium phosphate precipitation

WO 98/49293 - 9 - PCT/GB98/01206

of DNA is most commonly used (Bachetti et al., Proc. Natl. Acad. Sci. USA, 1977, 74, 1590-1594; Maitland et al., Cell, 1977, 14, 133-141), and represents a favoured technique in the context of the present invention.

A small percentage of the host cells takes up the recombinant DNA. In a small percentage of those, the DNA will integrate into the host cell chromosome. Because an antibitotic resistance marker gene, such as the neomycin or zeocin resistance gene, will have been incorporated into these host cells, they can be selected by isolating the individual clones which will grow in the presence of the chosen antibiotic, e.g. neomycin or zeocin. Each such clone may then tested to identify those which will produce the receptor. This may be achieved by inducing the production, for example with dexamethasone, and then detecting the presence of receptor by means of radioligand binding.

Alternatively, expression of the GABA receptor may be effected in Xenopus oocytes (see, for instance, Hadingham et al. Mol. Pharmacol., 1993, 44, 1211-1218). Briefly, isolated oocyte nuclei are injected directly with injection buffer or sterile water containing at least one alpha subunit, the theta subunit, and optionally one or more beta, gamma, delta or epsilon receptor subunits, engineered into a suitable expression vector. The oocytes are then incubated.

The expression of subunit combinations in the transfected oocytes may be demonstrated using conventional patch clamp assay. This assay measures the charge flow into and out of an electrode sealed on the surface of the cell. The flow of chloride ions entering the cell via the GABA gated ion channel is measured as a function of the current that leaves the cell to maintain electrical equilibrium within the cell as the gate opens.

In a further aspect, the present invention provides protein preparations of GABA receptor subunit combinations, especially human GABA receptor subunit combinations, derived from cultures of stably transfected eukaryotic cells.

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The protein preparations of GABA receptor subunit combinations can be recovered and purified from recombinant cell cultures by methods including ammonium sulfate or ethanol precipitation, acid extraction, anion or cation exchange chromatography, phosphocellulose chromatography, hydrophobic interaction chromatography, affinity chromatography, hydroxylapatite chromatography and lectin chromatography. Protein refolding steps can be used, as necessary, in completing configuration of the mature protein. Finally, high performance liquid chromatography (HPLC) can be employed for final purification steps.

The polypeptides of the present invention may be a naturally purified product, or a product of chemical synthetic procedures, or produced by recombinant techniques from a prokaryotic or eukaryotic host (for example, by bacterial, yeast, higher plant, insect and mammalian cells in culture). Depending upon the host employed in a recombinant production procedure, the polypeptides of the present invention may be glycosylated or may be non-glycosylated. Polypeptides of the invention may also include an initial methionine amino acid residue.

The GABA theta subunit polypeptide of the present invention is also useful for identifying other subunits of the GABA receptor. An example of a procedure for identifying these subunits comprises raising high titre polyclonal antisera against unique, bacterially expressed GABA theta polypeptides. These polyclonal antisera are then used to immunoprecipitate detergent-solubilized GABA receptors from a mammalian brain, for example, a rat brain.

The invention also provides preparations of membranes containing subunit combinations of the GABA receptor, especially human GABA receptor subunit combinations, derived from cultures of stably transfected eukaryotic cells.

The cell line, and the membrane preparations therefrom, according to the present invention have utility in screening and design of drugs

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WO 98/49293 - 11 - PCT/GB98/01206

which act upon the GABA receptor, for example benzodiazepines, barbiturates, β-carbolines and neurosteroids.

Receptor localisation studies using in situ hybridization in monkey brains shows that the θ subunit has a restricted localisation; residing mainly in components of the limbic system (involved in emotions such as rage, fear, motivation sexual behaviours and feeding); medial septum, cingulate cortex, the amygdala and hippocampal fields, in various hypothalamic nuclei, and in regions that have been associated with pain perception; the cingulate cortex, insular cortex, and in mid brain and pons structures.

The present invention accordingly provides the use of stably cotransfected cell lines described above, and membrane preparations derived therefrom, in screening for and designing medicaments which act upon GABA receptors comprising the θ subunit. Of particular interest in this context are molecules capable of interacting selectively with GABA receptors made up of varying subunit combinations. As will be readily apparent, the cell line in accordance with the present invention, and the membrane preparations derived therefrom, provide ideal systems for the study of structure, pharmacology and function of the various GABA receptor subtypes. In particular, preferred screens are functional assays utilizing the pharmacological properties of the GABA receptor subunit combinations of the present invention.

Thus, according to a further aspect of the present invention, there is provided a method for determining whether a ligand, not known to be capable of binding to a human GABAA receptor comprising the theta subunit, can bind to a human GABAA receptor comprising the theta subunit, which comprises contacting a mammalian cell comprising DNA molecules encoding at least one alpha receptor subunit, the theta receptor subunit, and optionally one or more beta, gamma, delta or epsilon receptor subunits with the ligand under conditions permitting binding of ligands known to bind to the GABAA receptor, detecting the presence of any of the

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ligand bound to the GABAA receptor comprising the theta subunit, and thereby determining whether the ligand binds to the GABAA receptor comprising the theta subunit. The theta subunit-encoding DNA in the cell may have a coding sequence substantially the same as the coding sequence shown in Figure 1 or Figure 2. Preferably, the mammalian cell is non-neuronal in origin. An example of a non-neuronal mammalian cell is a fibroblast cell such as an Ltk- cell. The preferred method for determining whether a ligand is capable of binding to a human $GABA_A$ receptor comprising the theta subunit comprises contacting a transfected non-neuronal mammalian cell (i.e. a cell that does not naturally express any type of GABAA receptor, and thus will only express such a receptor if it is transfected into the cell) expressing a GABAA receptor comprising the theta subunit on its surface, or contacting a membrane preparation from such a transfected cell, with the ligand under conditions which are known to prevail, and thus to be associated with, in vivo binding of the ligands to a GABAA receptor comprising the theta subunit, detecting the presence of any of the ligand being tested bound to the GABAA receptor comprising the theta subunit on the surface of the cell, and thereby determining whether the ligand binds to a human GABAA receptor comprising the theta subunit. This response system may be based on ion flux changes measured, for example, by scintillation counting (where the ion is radiolabelled) or by interaction of the ion with a fluorescent marker. Particularly suitable ions are chloride ions. Such a host system is conveniently isolated from pre-existing cell lines. Such a transfection system provides a complete response system for investigation or assay of the activity of human GABAA receptors comprising the theta subunit with ligands as described above. Transfection systems are useful as living cell cultures for competitive binding assays between known or candidate drugs and ligands which bind to the receptor and which are labeled by radioactive, spectroscopic or other reagents. Membrane preparations containing the receptor isolated from transfected cells are also useful for

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WO 98/49293 - 13 - PCT/GB98/01206

these competitive binding assays. A transfection system constitutes a "drug discovery system" useful for the identification of natural or synthetic compounds with potential for drug development that can be further modified or used directly as therapeutic compounds to activate, inhibit or modulate the natural functions of human GABAA receptors comprising the theta subunit. The transfection system is also useful for determining the affinity and efficacy of known drugs at human GABAA receptor sites comprising the theta subunit.

This invention also provides a method of screening drugs to identify drugs which specifically interact with, and bind to, a human GABAA receptor comprising the theta subunit on the surface of a cell which comprises contacting a mammalian cell comprising DNA molecules encoding at least one alpha receptor subunit, the theta receptor subunit and optionally one or more beta, gamma, delta or epsilon receptor subunits on the surface of a cell with a plurality of drugs, determining those drugs which bind to the mammalian cell, and thereby identifying drugs which specifically interact with, and bind to, human GABAA receptors comprising the theta subunit. The theta subunit-encoding DNA in the cell may have a coding sequence substantially the same as the coding sequence shown in Figure 1 or Figure 2. Preferably, the mammalian cell is non-neuronal in origin. An example of a non-neuronal mammalian cell is a fibroblast cell such as an Ltk- cell. Drug candidates are identified by choosing chemical compounds which bind with high affinity to the expressed GABAA receptor protein in transfected cells. using radioligand binding methods well known in the art. Drug candidates are also screened for selectivity by identifying compounds which bind with high affinity to one particular GABAA receptor combination but do not bind with high affinity to any other GABAA receptor combination or to any other known receptor site. Because selective, high affinity compounds interact primarily with the target GABAA receptor site after administration to the patient, the chances of

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producing a drug with unwanted side effects are minimized by this approach.

In the above screens, the mammalian cell may, for example, comprise DNA molecules encoding at least one alpha receptor subunit, the theta subunit, and optionally one or more gamma receptor subunits and optionally one or more beta receptor subunits.

More preferably, in the above screens, the mammalian cell comprises DNA molecules encoding at least one alpha receptor subunit, at least one gamma receptor subunit and the theta receptor subunit.

Ligands or drug candidates identified above may be agonists or antagonists at human GABAA receptors comprising the theta subunit, or may be agents which allosterically modulate a human GABAA receptor comprising the theta subunit. These ligands or drug candidates identified above may be employed as therapeutic agents, for example, for the modulation of emotions such as rage and fear, of sexual and appetite behaviours and of pain perception.

The ligands or drug candidates of the present invention thus identified as therapeutic agents may be employed in combination with a suitable pharmaceutical carrier. Such compositions comprise a therapeutically effective amount of the agonist or antagonist, and a pharmaceutically acceptable carrier or excipient.

Preferably the compositions containing the ligand or drug candidate identified according to the methods of the present invention are in unit dosage forms such as tablets, pills, capsules, wafers and the like.

Additionally, the therapeutic agent may be presented as granules or powders for extemporaneous formulation as volume defined solutions or suspensions. Alternatively, the therapeutic agent may be presented in ready-prepared volume defined solutions or suspensions. Preferred forms are tablets and capsules.

For preparing solid compositions such as tablets, the principal active ingredient is mixed with a pharmaceutical carrier, e.g. conventional

WO 98/49293 - 15 - PCT/GB98/01206

tableting ingredients such as corn starch, lactose, sucrose, sorbitol, talc, stearic acid, magnesium stearate, dicalcium phosphate or gums, and other pharmaceutical diluents, e.g. water, to form a solid preformulation composition containing a homogeneous mixture of a compound of the present invention, or a non-toxic pharmaceutically acceptable salt thereof. When referring to these preformulation compositions as homogeneous, it is meant that the active ingredient is dispersed evenly throughout the composition so that the composition may be readily subdivided into equally effective unit dosage forms such as tablets, pills and capsules. This solid preformulation composition is then subdivided into unit dosage forms of the type described above containing from 0.1 to about 500 mg of the active ingredient of the present invention. The tablets or pills of the novel composition can be coated or otherwise compounded to provide a dosage form affording the advantage of prolonged action. For example, the tablet or pill can comprise an inner dosage and an outer dosage component, the latter being in the form of an envelope over the former. The two components can be separated by an enteric layer which serves to resist disintegration in the stomach and permits the inner component to pass intact into the duodenum or to be delayed in release. A variety of materials can be used for such enteric layers or coatings, such materials including a number of polymeric acids and mixtures of polymeric acids with such materials as shellac, cetyl alcohol and cellulose acetate.

The liquid forms in which the novel compositions of the present invention may be incorporated for administration orally include aqueous solutions, suitably flavoured syrups, aqueous or oil suspensions, and flavoured emulsions with edible oils such as cottonseed oil, sesame oil, coconut oil, peanut oil or soybean oil, as well as elixirs and similar pharmaceutical vehicles. Suitable dispersing or suspending agents for aqueous suspensions include synthetic and natural gums such as tragacanth, acacia, alginate, dextran, sodium carboxymethylcellulose, methylcellulose, polyvinyl-pyrrolidone or gelatin.

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Compositions of the present invention may also be administered via the buccal cavity using conventional technology, for example, absorption wafers.

Compositions in the form of tablets, pills, capsules or wafers for oral administration are particularly preferred.

A minimum dosage level for the ligand or drug candidate identified according to the methods of the present invention is about 0.05mg per day, preferably about 0.5mg per day and especially about 2.5mg per day. A maximum dosage level for the ligand or drug candidate is about 3000mg per day, preferably about 1500mg per day and especially about 500mg per day. The compounds are administered on a regimen of 1 to 4 times daily, preferably once or twice daily, and especially once a day.

It will be appreciated that the amount of the therapeutic agent required for use therapy will vary not only with the particular compounds or compositions selected but also with the route of administration, the nature of the condition being treated, and the age and condition of the patient, and will ultimately be at the discretion of the patient's physician or pharmacist.

20 <u>DESCRIPTION OF FIGURES</u>

Figure 1: Nucleotide sequence for the theta subunit, together with its deduced amino acid sequence corresponding thereto (SEQ.ID.NO.1 and SEQ.ID.NO.2, respectively)

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Figure 2: Preferred nucleotide sequence for the theta subunit, together with its deduced amino acid sequence corresponding thereto (SEQ.ID.NO.3 and SEQ.ID.NO.4, respectively).

Figure 3: GABA dose-response curves on HEK cells transiently transfected with and without θ subunit-containing GABA-A receptors ($\alpha_2\beta_1\theta\gamma_1$ and $\alpha_2\beta_1\gamma_1$).

The following non-limiting Examples illustrate the present invention.

EXAMPLE 1

10 ISOLATION AND SEQUENCING OF A cDNA ENCODING THE HUMAN GABAA RECEPTOR θ SUBUNIT.

The Genbank database was searched with GABAA receptor polypeptide amino acid sequences using the BLAST searching algorithm, and a number of homologous sequences identified. One of these U47334 was investigated in more detail. U47334 contained sequences homologous to part of the amino-terminal extracellular domain and the TM4 spanning domain of other GABAA receptor subunits, but did not appear to contain any sequence homologous to the regions spanning these domains. 20 Polymerase chain reaction (PCR) was performed to determine if the size if the U47334 sequence was correct, or was for example, the result of an incorrect splicing event. For PCR, a sense (5' gcaaatgaagctgtggttc 3') (SEQ.ID.NO. 5) and antisense (5' caatgttgaacaacccaaag 3') (SEQ.ID.NO. 6) primer were generated from the U47334 sequence, and PCR performed 25 using standard conditions (Whiting et al, PNAS) using human whole brain cDNA (Clontech) as a template. A second PCR reation was then performed using nested sense (5' gcctgagaccgaattttgg 3') (SEQ.ID.NO. 7) and antisense (5' ggaaccgggaccacttgtc 3') (SEQ.ID.NO. 8) primers generated from the U47334 sequence, and using the products from the 30 first PCR as a template. A single PCR product of approximately 1600 bp was obtained suggesting that the U47334 sequence represents an

incorrectly processed message. This product was sequenced directly using an Applied Biosystems 373 DNA sequencer and dye terminator chemistry.

cDNA sequences 5' and 3' of the U47334 sequence were obtained by 5'- and 3'-anchored PCR using human brain Marathon cDNA cloning kit (Clontech) according to the manufacturer's protocols. The nested antisense (5' tagtccagggtcaagttc 3' and 5' tagtatgctaagcgtgaatc 3') (SEQ.ID.NOS. 9 and 10) and sense (5' gagtttgaggatagttgc 3' and 5' tgctccttcactgaaggg 3') (SEQ.ID.NOS. 11 and 12) primers were derived from both the U47334 sequence and the sequence from the initial PCR amplifications. The PCR products were sequenced directly as previously described.

A full length cDNA was generated by PCR using primers derived from sequences downstream of the innitiating ATG (5' ccatgactcaagcttgccaccatgctgcgagccgcagtgatc 3', incorporating a HindIII site) (SEQ.ID.NO. 13) and in the 3' UT of the anchored PCR product (5' tgaaaggagcacagcacagtgctcccg 3') (SEQ.ID.NO. 14). The PCR product (1958 bp) was cloned into pMOS (Amersham), subcloned into pCDNAI Amp (Invitrogen), and sequenced completey on both strands by primer walking. Sequence analysis was performed using Inherit (Applied Biosystems), Sequencher (Genecodes), and Genetics Computer Group (Univ. Wisconsin) computer programs.

The coding region encodes 627 amino acids and has all the structural motifs expected of a ligand gated ion channel subunit. Comparison with other ligand gated ion channel subunits indicates that it is most similar to GABAA receptor subunits, the highest homology being with the β_1 subunit (45 % identity). However, this sequence identity is sufficiently low as to indicate that the new subunit cannot be classified as a fourth human β subunit, but represents a novel class of subunit, classified as θ , within the GABA receptor gene family.

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EXAMPLE 2

LOCALISATION OF THE θ SUBUNIT IN MONKEY BRAIN BY *IN* SITU HYBRIDISATION.

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Antisense oligonucleotide probes to the human θ subunit sequence were generated on an Applied Biosystems Automated DNA synthesiser Probe 1

5' CTG-CTT-CTT-GCA-CAC-CCT-TCT-CGC-CAT-GGT-GAA-GCA-TGG-GCT-TCC 3' (SEQ.ID.NO. 15)

Probe 2

5'TGT-CGC-CTA-GGC-TGG-CGC-CGA-GGT-CCT-CGA-CTG-TAG-AAA-AGA-TAG 3' (SEQ.ID.NO. 16)

Each oligonucleotide was 3'-end labelled with [35S] deoxyadenosine 5'-(thiotriphosphate) in a 30:1 molar ratio of 35 S-isotope:oligonucleotide using terminal deoxynucleotidyl transferase for 15 min at 37°C in the reaction buffer supplied. Radiolabelled oligonucleotide was separated from unincorporated nucleotides using Sephadex G50 spin columns. The specific activities of the labelled probes in several labelling reactions varied from 1.2-2.3 x 10⁹ cpm/mg. Monkey brains were removed and fresh frozen in 1 cm blocks. 12 µm sections were taken and fixed for in situ hybridisation. Hybridisation of the sections was carried out according to the method of Sirinathsingji and Dunnett (Imaging gene expression in neural graft; Molecular Imaging in Neuroscience: A Practical Approach, N.A. Sharif (ed), Oxford University Press, Oxford, pp43-70, 1993). Briefly, sections were removed from alcohol, air dried and 3 x10⁵ cpm of each ³⁵S-labelled probe in 100μl of hybridisation buffer was applied to each slide. Labelled "antisense" probe was also used in the presence of an excess (100x) concentration of unlabelled antisense probe to define nonspecific hybridisation. Parafilm coverslips were placed over the sections

which were incubated overnight (about 16 hr) at 37°C. Following hybridisation the sections were washed for 1 hr at 57°C in 1xSSC then rinsed briefly in 0.1 x SSC, dehydrated in a series of alcohols, air dried and exposed to Amersham Hyperfilm β max X-ray film and the relative distribution of the mRNA assessed for a variety of brain regions.

Messenger RNA for the subunit was seen in components of the limbic system (involved in emotions such as rage, fear, motivation sexual behaviours and feeding); medial septum, cingulate cortex, the amygdala and hippocampal fields (dentate gyrus, CA3, CA2, CA1) and in various hypothalamic nuclei (often associated with the limbic system). Messenger RNA was also present in regions that have been associated with pain perception; the cingulate cortex, insular cortex, and in mid brain and pons structures (e.g. central grey and reticular formation).

EXAMPLE 3

LOCALISATION OF THE θ SUBUNIT IN MONKEY BRAIN BY WESTERN BLOT ANALYSIS AND IMMUNOCYTOCHEMISTRY

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Antibodies to the human GABAA Theta subunit were generated by sub-cutaneous injection of two New Zealand White rabbits with a glutathione-S-transferase (GST) fusion protein consisting of residues 353-595 of the large cytoplasmic loop region of the theta subunit. DNA encoding this region was cloned into the bacterial expression vector pGEX-2T (Pharmacia), transformed into *E. coli* DH10B cells (Life Technologies), and expression of the fusion protein was carried out using the Pharmacia protocols. The bacterial cells were incubated on ice in STE solution (150 mM NaCl, 10 mM Tris-HCl pH 8, 1 mM EDTA) containing 100 µg/ml Lysozyme for 20 min before the addition of N-lauryl sarkosine to 1.5 % (w/v). The bacterial slurry was sonicated on ice, and any insoluble matter

WO 98/49293 - 21 - PCT/GB98/01206

removed by centrifugation. Triton X-100 was added to 3 % (v/v) final and the GST-fusion protein purified by glutathione-agarose affinity chromatography. Columns were washed extensively with PBS and the bound protein eluted with 20 mM free glutathione in 150 mM NaCl, 100 mM Tris-HCl pH 9, 1 mM EDTA, 1 mM Dithiothreitol. Eluted protein was concentrated by precipitation with 5 volumes of cold acetone, resuspended in water, and stored at -70 °C until use.

For western blot analysis tissue samples were removed and dissected out on a glass plate at 4°C. The tissue was homogenised in 50mM Tris, pH 7.5, containing 1mM PMSF, 1µM pepstatin A. The homogenate was centrifuged (2000 X g) for 10 minutes and the supernatant was centrifuged at 20,000 X g for 45 minutes. The pellet was resuspended in 50mM Tris and recentrifuged. The final pellet was resuspended in 50mM Tris pH 7.4 containing protease inhibitors and detergent (Na-deoxycholate:0.25%, 150mM NaCl, 1mM EDTA, 1mM PMSF, 1µM pepstatin and leupeptin. Membrane preparations were separated on a 10 % Tris tricine polyacrylamide gel and electrophoretically transferred to nitrocellulose. Nitrocellulose was blocked with 5% non-fat milk (marvel™)/PBS/Tween (0.5%) for 1 hour at room temperature. The anti θ subunit antibody was used at a concentration of 1:500 made up in PBS/Tween/milk at 4°C overnight, washed and then incubated with anti-rabbit IgG HRP linked (Amersham) at 1:1000 in PBS/Tween/milk for one hour at room temperature. The filters were washed, incubated in ECL (Amersham) for 1min and opposed to film. A single band of approximately 60-66kDa was visualised in brainstem and striatal membranes, close to the predicted molecular weight for the θ subunit of 68-74 kDa.

For localisation of the θ subunit by immunocytochemistry a rhesus monkey was deeply anesthetised with ketamine and sodium

30 pentobarbitone and transcardially perfused with saline, followed by 10% formal saline. The brain was removed, post fixed for 24 hours, and sliced

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into coronal blocks, which were then dehydrated through graded alcohols, cleared and embedded in paraffin wax. Coronal sections (8µm) were cut on a base sledge microtome and mounted on glass microscope slides. Sections were deparaffinised, rehydrated and rinsed in 0.1M phosphate buffered saline (PBS). In order to enhance the immunoreactivity sections were subjected to antigen retrieval techniques. Briefly, sections were placed in 0.1M citrate buffer pH 6.0 and given two 5 minute bursts at full power in a conventional microwave oven (800W). Once rinsed in PBS, sections were incubated in 5% normal goat serum in PBS, for 1 hr to block background staining. Sections were then incubated overnight at +4°C in the anti θ subunit rabbit polyclonal antibody (1:1000 diluted in blocking buffer). Immunoreactivity was visualised using the Vector elite™ system (Vector Laboratories, Peterborough, U.K.), followed by development in diaminobenzidine (DAB) (Sigma, U.K.). Sections were counterstained in Gill's haematoxylin (Biomen, High Wycombe, U.K.), dehydrated and mounted for microscopical examination. For comparison, samples of 10% formalin immersion fixed post mortem human brainstem were processed in an identical manner. Comparable sections were used to detect θ subunit and tyrosine hydroxylase (Institut Jacques Boy, Reims, France) immunoreactivity by the application of 35S-labeled goat anti rabbit immunoglobulin 1:100 (Amersham Life Sciences, U.K.) for 1 hr. Slides were rinsed in distilled water, dehydrated to 95% ethanol, air dried and exposed to Amersham Hyperfilm βmax. Sections used for the immunofluorescent colocalisation of θ subunit and tyrosine hydroxylase were pretreated in the same manner, anti θ subunit immunoreactivity was detected using firstly a biotinylated anti rabbit ;1:200 (Vector Laboratories) followed by FITC conjugated streptavidin (Sigma, U.K.). The second rabbit polyclonal serum, anti tyrosine hydroxylase, was again visualised using biotinylated anti rabbit, reacted with Cy3 conjugated strepavidin (Sigma, U.K.). Sections were counterstained with Hoescht

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WO 98/49293 - 23 - PCT/GB98/01206

33258 (0.5µg/ml). To avoid any crossreactivity of the detection systems, sections were placed in boiling distilled water for 5 minutes prior to the application of the second primary antibody and its subsequent detection.. The distribution of the θ subunit immunoreactivity in monkey brain reflected the distribution of the θ mRNA observed by in situ hybridisation studies (Example 2). Labelled neurons were observed of hypothalamic and cortical pyramidal neurones. Significant labellingwas observed of cells in the brainstem, including the substantia nigra pars compacta, ventral and lateral tegmental areas, pigmented neurones of the locus coeruleus and restricted population within the dorsal raphe. Labelling of cell terminals within the caudate putamen was also observed. This distribution was found to closely resemble the distribution of tyrosine hydroxylase immunoreactivity, a marker of catocholaminergic neurones and their processes, visualised by immunoautoradiography. θ subunit colocalisation with tyrosine hydroxylase containing neurons was confirmed, using combination immunofluorescence. The expression of the θ subunit seen in both the catocholaminergic neurons of the substantia nigra pars compacta and locus coeruleus was further substantiated in sections of human post mortem brainstem.

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EXAMPLE 4

CONSTRUCTION OF AN LTK- CELL LINE EXPRESSING THE THETA RECEPTOR SUBUNIT

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A chimeric construct of the theta subunit was constructed in the mammalian expression vector pcDNA3.1Zeo (Invitrogen) that consisted of bases -224 to +99 of bovine GABA_A α 1 gene, a sequence encoding the c-myc epitope tag (residues 410-419 of the human oncogene product c-myc), a cloning site encoding the amino acids aparagine - serine - glycine, and

DNA encoding residues 22-627 of the GABAA θ gene product. This construct was linearised and the DNA transfected into a clonal population of mouse Ltk- cells that had previously been shown to be stably transfected with the GABAA receptor subunits $\alpha_2\beta_1\gamma_1$ and separately an Ltk- line stably transfected with $\alpha_2\beta_3\gamma_2$. The resultant cells were clonally selected with Zeocin selection (100 µg/ml), and screened to verify stable intrgration and expression of $\alpha_2\beta_1\theta\gamma_1$ and $\alpha_2\beta_3\theta\gamma_2$ respectively.

EXAMPLE 5

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WHOLE CELL PATCH-CLAMP OF HEK 293 CELLS
TRANSIENTLY TRANSFECTED WITH HUMAN GABA-A
RECEPTORS

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Experiments were performed on HEK 293 cells transiently transfected with human cDNA combinations $\alpha 2\beta 1\gamma 1,$ and $~\alpha 2\beta 1\theta \gamma 1$ (4µgs of cDNA total per cover-slip) using calcium phosphate precipitation (Chen and Okayama, 1988) as previously described (Hadingham et al, 1993). Glass cover-slips containing the cells in a monolayer culture were transferred to a perspex chamber on the stage of Nikon Diaphot inverted microscope. Cells were continuously perfused with a solution containing 124mM NaCl, 2mM KCl, 2mM CaCl $_2$, 1mM MgCl $_2$, 1.25mM KH $_2$ PO $_4$, 25mM NaHCO3, 11mM D-glucose, at pH 7.2, and observed using phasecontrast optics. Patch-pipettes were pulled with an approximate tip diameter of $2\mu m$ and a resistance of $4M\Omega$ with borosilicate glass and filled with 130mM CsCl, 10mM HEPES, 10mM EGTA, 3mM Mg+-ATP, pH adjusted to 7.3 with CsOH. Cells were patch-clamped in whole-cell mode using an Axopatch 200B patch-clamp amplifier. Drug solutions were applied by a double-barrelled pipette assembly, controlled by a stepping motor attached to a Prior manipulator, enabling rapid equilibration

around the cell. Increasing GABA concentrations were applied for 2sec pulses with a 30sec interval between applications. Non-cumulative concentration-response curves to GABA were constructed. Curves were fitted using a non-linear square-fitting program to the equation $f(x) = B_{\text{MAX}}/[1+(\text{EC}_{50}/x)^n]$ where x is the drug concentration, EC₅₀ is the concentration of drug eliciting a half-maximal response and n is the Hill coefficient. EC₅₀'s were analysed by unpaired students t-test.

The GABA EC₅₀ of HEK 293 cells transiently expressing the GABA_A receptor subunit combination $\alpha_2\beta_1\theta\gamma_1$ is significantly lower than that of HEK 293 cells transiently expressing the GABA_A receptor subunit combination $\alpha_2\beta_1\gamma_1$ (see Figure 3).

	$\alpha_2\beta_1\gamma_1$	$\alpha_2\beta_1\theta\gamma_1$				
EC_{50}	16.7±3.7 nM	62.7±6.7 nM*				
Slope	1.6±0.2	1.5±0.1				

* p<0.001

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT:
 - (A) NAME: Merck Sharp & Dohme Limited
 - (B) STREET: Terlings Park, Eastwick Road
 - (C) CITY: Harlow
 - (D) STATE: Essex
 - (E) COUNTRY: England
 - (F) POSTAL CODE (ZIP): CM20 2QR
 - (G) TELEPHONE: +44 1279 440175
 - (H) TELEFAX: +44 1279 440717
- (ii) TITLE OF INVENTION: Human theta subunit of the GABA-A receptor
- (iii) NUMBER OF SEQUENCES: 16
- (iv) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.30 (EPO)
- (2) INFORMATION FOR SEQ ID NO: 1:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1884 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: cDNA
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:
- ATGCTGCGAG CCGCAGTGAT CCTGCTGCTC ATCAGGACCT GGCTCGCGGA GGGCAACTAC 60
 CCCAGTCCCA TCCCGAAATT CCACTTCGAG TTCTCCTCTG CTGTGCCCGA AGTCGTCCTG 120
- AACCTCTTCA ACTGCAAAAA TTGTGCAAAT GAAGCTGTGG TTCAAAAGAT TTTGGACAGG 180
- GTGCTGTCAA GATACGATGT CCGCCTGAGA CCGAATTTTG GAGGTGCCCC TGTGCCTGTG 240
- AGAATATCTA TTTATGTCAC GAGCATTGAA CAGATCTCAG AAATGAATAT GGACTACACG 300
- ATCACGATGT TTTTTCATCA GACTTGGAAA GATTCACGCT TAGCATACTA TGAGACCACC 360

CTGAACTTGA	CCCTGGACTA	TCGGATGCAT	GAGAAGTTGT	GGGTCCCTGA	CTGCTACTTT	420
CTGAACAGCA	AGGATGCTTT	CGTGCATGAT	GTGACTGTGG	AGAATCGCGT	GTTTCAGCTT	480
CACCCAGATG	GAACGGTGCG	GTACGGCATC	CGACTCACCA	CTACAGCAGT	TTGTTCCCTG	540
GATCTGCATA	AATTCCCTAT	GGACAAGCAG	GCCTGCAACC	TGGTGGTAGA	GAGCTATGGT	600
TACACGGTTG	AAGACATCAT	ATTATTCTGG	GATGACAATG	GGAACGCCAT	CCACATGACT	660
GAGGAGCTGC	ATATCCCTCA	GTTCACTTTC	CTGGGAAGGA	CGATTACTAG	CAAGGAGGTG	720
TATTTCTACA	CAGGTTCCTA	CATACGCCTG	ATACTGAAGT	TCCAGGTTCA	GAGGGAAGTT	780
AACAGCTACC	TTGTGCAAGT	CTACTGGCCT	ACTGTCCTCA	CCACTATTAC	CTCTTGGATA	840
TCGTTTTGGA	TGAACTATGA	TTCCTCTGCA	GCCAGGGTGA	CAATTGGCTT	AACTTCAATG	900
CTCATCCTGA	CCACCATCGA	CTCACATCTG	CGGGATAAGC	TCCCCAACAT	TTCCTGTATC	960
AAGGCCATTG	ATATCTATAT	CCTCGTGTGC	TTGTTCTTTG	TGTTCCTGTC	CTTGCTGGAG	1020
TATGTCTACA	TCAACTATCT	TTTCTACAGT	CGAGGACCTC	GGCGCCAGCC	TAGGCGACGC	1080
AGGAGACCCC	GAAGAGTCAT	TGCCCGCTAC	CGCTACCAGC	AAGTGGTGGT	AGGAAACGTG	1140
CAGGATGGCC	TGATTAACGT	GGAAGACGGA	GTCAGCTCTC	TCCCCATCAC	CCCAGCGCAG	1200
GCCCCCTGG	CAAGCCCGGA	AAGCCTCGGT	TCTTTGACGT	CCACCTCCGA	GCAGGCCCAG	1260
CTGGCCACCT	CGGAAAGCCT	CAGCCCACTC	ACTTCTCTCT	CAGGCCAGGC	CCCCTGGCC	1320
ACTGGAGAAA	GCCTGAGCGA	TCTCCCCTCC	ACCTCAGAGC	AGGCCCGGCA	CAGCTATGGT	1380
GTTCGCTTTA	ATGGTTTCCA	GGCTGATGAC	AGTATTATTC	CTACCGAAAT	CCGCAACCGT	1440
GTCGAAGCCC	ATGGCCATGG	TGTTACCCAT	GACCATGAAG	ATTCCAATGA	GAGCTTGAGC	1500
TCGGATGAGC	GCCATGGCCA	TGGCCCCAGT	GGGAAGCCCA	TGCTTCACCA	TGGCGAGAAG	1560
GGTGTGCAAG	AAGCAGGCTG	GGACCTTGAT	GACAACAATG	ACAAGAGCGA	CTGCCTTGCC	1620
ATTAAGGAGC	AATTCAAGTG	TGATACTAAC	AGTACCTGGG	GCCTTAATGA	TGATGAGCTC	1680
GTGGCCCATG	GCCAAGAGAA	GGACAGTAGC	TCAGAGTCTG	AGGATAGTTG	CCCCCCAAGC	1740
CCTGGGTGCT	CCTTCACTGA	AGGGTTCTCC	TTCGATCTCT	TTAATCCTGA	CTACGTCCCA	1800
AAGGTCGACA	AGTGGTCCCG	GTTCCTCTTC	CCTCTGGCCT	TTGGGTTGTT	CAACATTGTT	1860
TACTGGGTAT	ACCATATGTA	TTAG				1884

(2) INFORMATION FOR SEQ ID NO: 2:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 627 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

Met Leu Arg Ala Ala Val Ile Leu Leu Leu Ile Arg Thr Trp Leu Ala Glu Gly Asn Tyr Pro Ser Pro Ile Pro Lys Phe His Phe Glu Phe Ser Ser Ala Val Pro Glu Val Val Leu Asn Leu Phe Asn Cys Lys Asn Cys Ala Asn Glu Ala Val Val Gln Lys Ile Leu Asp Arg Val Leu Ser Arg Tyr Asp Val Arg Leu Arg Pro Asn Phe Gly Gly Ala Pro Val Pro Val Arg Ile Ser Ile Tyr Val Thr Ser Ile Glu Gln Ile Ser Glu Met Asn 85 90 Met Asp Tyr Thr Ile Thr Met Phe Phe His Gln Thr Trp Lys Asp Ser 100 105 Arg Leu Ala Tyr Tyr Glu Thr Thr Leu Asn Leu Thr Leu Asp Tyr Arg 120 Met His Glu Lys Leu Trp Val Pro Asp Cys Tyr Phe Leu Asn Ser Lys 130 Asp Ala Phe Val His Asp Val Thr Val Glu Asn Arg Val Phe Gln Leu His Pro Asp Gly Thr Val Arg Tyr Gly Ile Arg Leu Thr Thr Ala 165 170 Val Cys Ser Leu Asp Leu His Lys Phe Pro Met Asp Lys Gln Ala Cys 180 Asn Leu Val Val Glu Ser Tyr Gly Tyr Thr Val Glu Asp Ile Ile Leu 200 Phe Trp Asp Asp Asn Gly Asn Ala Ile His Met Thr Glu Glu Leu His 210 Ile Pro Gln Phe Thr Phe Leu Gly Arg Thr Ile Thr Ser Lys Glu Val 225 230

	Tyr	Phe	Tyr	Thr	Gly 245	Ser	Tyr	Ile	Arg	Leu 250	Ile	Leu	Lys	Phe	Gln 255	Val
	Gln	Arg	Glu	Val 260	Asn	Ser	Tyr	Leu	Val 265	Gln	Val	Tyr	Trp	Pro 270	Thr	Val
	Leu			Ile										Tyr	Asp	Ser
	Ser	Ala 290	Ala	Arg	Val	Thr	Ile 295	Gly	Leu	Thr	Ser	Met 300	Leu	Ile	Leu	Thr
	Thr 305	Ile	Asp	Ser	His	Leu 310	Arg	Àsp	Lys	Leu	Pro 315	Asn	Ile	Ser	Cys	Ile 320
	Lys	Ala	Ile	Asp	Ile 325	Tyr	Ile	Leu	Val	Cys 330	Leu	Phe	Phe	Val	Phe 335	Leu
	Ser	Leu	Leu	Glu 340	Tyr	Val	Tyr	Ile	Asn 345	Tyr	Leu	Phe	Tyr	Ser 350	Arg	Gly
	Pro	Arg	Arg 355	Gl'n	Pro	Arg	Arg	Arg 360	Arg	Arg	Pro	Arg	Arg 365	Val	Ile	Ala
	Arg	Tyr 370	Arg	Tyr	Gln	Gln	Val 375	Val	Val	Gly	Asn	Val 380	Gln	Asp	Gly	Leu
	Ile 385	Asn	Val	Glu	Asp	Gly 390	Val	Ser	Ser	Leu	Pro 395	Ile	Thr	Pro	Ala	Gln 400
	Ala	Pro	Leu	Ala	Ser 405	Pro	Glu	Ser	Leu	Gly 410	Ser	Leu	Thr	Ser	Thr 415	Ser
	Glu	Gln	Ala	Gln 420	Leu	Ala	Thr	Ser	Glu 425	Ser	Leu	Ser	Pro	Leu 430	Thr	Ser
	Leu	Ser	Gly 435	Gln	Ala	Pro	Leu	Ala 440	Thr	Gly	Glu	Ser	Leu 445	Ser	Asp	Leu
	Pro	Ser 450	Thr	Ser	Glu	Gln	Ala 455	Arg	His	Ser	Tyr	Gly 460	Val	Arg	Phe	Asn
-	Gly 465	Phe	Gĺn	Ala	Asp	Asp 470	Ser	Ile	Ile	Pro	Thr 475	Glu	Ile	Arg	Asn	Arg 480
	Val	Glu	Ala	His	Gly 485	His	Gly	Val	Thr	His 490	Asp	His	Glu	Asp	Ser 495	Asn
	Glu	Ser	Leu	Ser 500	Ser	Asp	Glu	Arg	His 505	Gly	His	Gly	Pro	Ser 510	Gly	Lys
	Pro	Met	Leu 515	His	His	Gly	Glu	Lys 520	Gly	Val	Gln	Glu	Ala 525	Gly	Trp	Asp
	Leu	Asp 530	Asp	Asn	Asn	Asp	Lys 535	Ser	Asp	Cys	Leu	Ala 540	·Ile	Lys	Glu	Gln

Phe Lys Cys Asp Thr Asn Ser Thr Trp Gly Leu Asn Asp Asp Glu Leu Val Ala His Gly Gln Glu Lys Asp Ser Ser Ser Glu Ser Glu Asp Ser 570 Cys Pro Pro Ser Pro Gly Cys Ser Phe Thr Glu Gly Phe Ser Phe Asp

Leu Phe Asn Pro Asp Tyr Val Pro Lys Val Asp Lys Trp Ser Arg Phe

Leu Phe Pro Leu Ala Phe Gly Leu Phe Asn Ile Val Tyr Trp Val Tyr

His Met Tyr 625

(2) INFORMATION FOR SEQ ID NO: 3:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1884 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

ATGCTGCGAG CCGCAGTGAT CCTGCTGCTC ATCAGGACCT GGCTCGCGGA GGGCAACTAC CCCAGTCCCA TCCCGAAATT CCACTTCGAG TTCTCCTCTG CTGTGCCCGA AGTCGTCCTG 120 AACCTCTTCA ACTGCAAAAA TTGTGCAAAT GAAGCTGTGG TTCAAAAGAT TTTGGACAGG 180 GTGCTGTCAA GATACGATGT CCGCCTGAGA CCGAATTTTG GAGGTGCCCC TGTGCCTGTG 240 AGAATATCTA TTTATGTCAC GAGCATTGAA CAGATCTCAG AAATGAATAT GGACTACACG 300 ATCACGATGT TTTTTCATCA GACTTGGAAA GATTCACGCT TAGCATACTA TGAGACCACC 360 CTGAACTTGA CCCTGGACTA TCGGATGCAT GAGAAGTTGT GGGTCCCTGA CTGCTACTTT 420 TTGAACAGCA AGGATGCTTT CGTGCATGAT GTGACTGTGG AGAATCGCGT GTTTCAGCTT 480 CACCCAGATG GAACGGTGCG GTACGGCATC CGACTCACCA CTACAGCAGC TTGTTCCCTG 540 GATCTGCATA AATTCCCTAT GGACAAGCAG GCCTGCAACC TGGTGGTAGA GAGCTATGGT 600 TACACGGTTG AAGACATCAT ATTATTCTGG GATGACAATG GGAACGCCAT CCACATGACT 660

GAGGAGCTGC ATATCCCTCA GTTCACTTC CTGGGAAGGA CGATTACTAG CAAGGAGGTG 720 TATTTCTACA CAGGTTCCTA CATACGCCTG ATACTGAAGT TCCAGGTTCA GAGGGAAGTT 780 AACAGCTACC TTGTGCAAGT CTACTGGCCT ACTGTCCTCA CCACTATTAC CTCTTGGATA 840 TCGTTTTGGA TGAACTATGA TTCCTCTGCA GCCAGGGTGA CAATTGGCTT AACTTCAATG 900 CTCATCCTGA CCACCATCGA CTCACATCTG CGGGATAAGC TCCCCAACAT TTCCTGTATC 960 AAGGCCATTG ATATCTATAT CCTCGTGTGC TTGTTCTTTG TGTTCCTGTC CTTGCTGGAG 1020 TATGTCTACA TCAACTATCT TTTCTACAGT CGAGGACCTC GGCGCCAGCC TAGGCGACAC 1080 AGGAGACCCC GAAGAGTCAT TGCCCGCTAC CGCTACCAGC AAGTGGTGGT AGGAAACGTG 1140 CAGGATGGCC TGATTAACGT GGAAGACGGA GTCAGCTCTC TCCCCATCAC CCCAGCGCAG 1200 GCCCCCTGG CAAGCCCGGA AAGCCTCGGT TCTTTGACGT CCACCTCCGA GCAGGCCCAG 1260 ACTGGAGAAA GCCTGAGCGA TCTCCCCTCC ACCTCAGAGC AGGCCCGGCA CAGCTATGGT 1380 GTTCGCTTTA ATGGTTTCCA GGCTGATGAC AGTATTTTTC CTACCGAAAT CCGCAACCGT 1440 GTCGAAGCCC ATGGCCATGG TGTTACCCAT GACCATGAG ATTCCAATGA GAGCTTGAGC 1500 TCGGATGAGC GCCATGGCCA TGGCCCCAGT GGGAAGCCCA TGCTTCACCA TGGCGAGAAG 1560 GGTGTGCAAG AAGCAGGCTG GGACCTTGAT GACAACAATG ACAAGAGCGA CTGCCTTGCC 1620 ATTAAGGAGC AATTCAAGTG TGATACTAAC AGTACCTGGG GCCTTAATGA TGATGAGCTC 1680 ATGGCCCATG GCCAAGAGAA GGACAGTAGC TCAGAGTCTG AGGATAGTTG CCCCCCAAGC 1740 CCTGGGTGCT CCTTCACTGA AGGGTTCTCC TTCGATCTCT TTAATCCTGA CTACGTCCCA 1800 AAGGTCGACA AGTGGTCCCG GTTCCTCTTC CCTCTGGCCT TTGGGTTGTT CAACATTGTT 1860 TACTGGGTAT ACCATATGTA TTAG 1884

(2) INFORMATION FOR SEQ ID NO: 4:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 627 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

Met 1	: Le	u Arg	g Ala	a Ala 5	a Val	. Ile	e Let	ı Lei	Lei 10	u Ile	e Arg	g Thi	r Tr	p Let 15	ı Ala
Glı	ı Gly	y Ası	туі 20	Pro	Ser	Pro	Ile	25	b Lys	s Phe	e His	s Phe	Gl: 30	ı Phe	e Ser
Ser	Ala	a Val	Pro	Glu	ı Val	Val	Leu 40	ı Asr	Let	ı Phe	e Asr	Cys	. Lys	s Asr	ı Cys
Ala	Asr 50	ı Glu	ı Ala	. Val	Val	Gln 55	Lys	Ile	. Leu	ı Asp	Arg 60	, Val	Let	ı Ser	Arg
Tyr 65	Asp	Val	Arg	Leu	Arg 70	Pro	Asn	Phe	Gly	75	Ala	Pro	Val	Pro	Val 80
Arg	Ile	: Ser	Ile	Tyr 85	Val	Thr	Ser	Ile	Glu 90	Gln	Ile	Ser	Glu	Met 95	Asn
Met	Asp	Tyr	Thr 100	Ile	Thr	Met	Phe	Phe 105	His	Gln	Thr	Trp	Lys 110		Ser
Arg	Leu	Ala 115	Tyr	Tyr	Glu	Thr	Thr 120	Leu	Asn	Leu	Thr	Leu 125	Asp	Tyr	Arg
Met	His 130	Glu	Lys	Leu	Trp	Val 135	Pro	Asp	Cys	Tyr	Phe 140		Asn	Ser	Lys
Asp 145	Ala	Phe	Val	His	Asp 150	Val	Thr	Val	Glu	Asn 155	Arg	Val	Phe	Gln	Leu 160
His	Pro	Asp	Gly	Thr 165	Val	Arg	Tyr	Gly	Ile 170	Arg	Leu	Thr	Thr	Thr 175	Ala
Ala	Cys	Ser	Leu 180	Asp	Leu	His	Lys	Phe 185	Pro	Met	Asp	Lys	Gln 190	Ala	Cys
		195		Glu			200					205			
Phe	Trp 210	Asp	Asp	Asn	Gly	Asn 215	Ala	Ile	His	Met	Thr 220	Glu	Glu	Leu	His
Ile 225	Pro	Gln	Phe	Thr	Phe 230	Leu	Gly	Arg _.	Thr	Ile 235	Thr	Ser	Lys	Glu	Val -
Tyr	Phe	Tyr	Thr	Gly 245	Ser	Tyr	Ile	Arg	Leu 250	Ile	Leu	Lys	Phe	Gln 255	Val
Gln	Arg	Glu	Val 260	Asn	Ser	Tyr	Leu	Val 265	Gln	Val	Tyr	Trp	Pro 270	Thr	Val
Leu	Thr	Thr 275	Ile	Thr	Ser	Trp	Ile 280	Ser	Phe	Trp	Met	Asn 285	Tyr	Asp	Ser

Ser Ala Ala Arg Val Thr Ile Gly Leu Thr Ser Met Leu Ile Leu Thr 295 300 Thr Ile Asp Ser His Leu Arg Asp Lys Leu Pro Asn Ile Ser Cys Ile 315 Lys Ala Ile Asp Ile Tyr Ile Leu Val Cys Leu Phe Phe Val Phe Leu 325 330 Ser Leu Leu Glu Tyr Val Tyr Ile Asn Tyr Leu Phe Tyr Ser Arg Gly Pro Arg Arg Gln Pro Arg Arg His Arg Arg Pro Arg Arg Val Ile Ala Arg Tyr Arg Tyr Gln Gln Val Val Gly Asn Val Gln Asp Gly Leu 375 Ile Asn Val Glu Asp Gly Val Ser Ser Leu Pro Ile Thr Pro Ala Gln 390 395 Ala Pro Leu Ala Ser Pro Glu Ser Leu Gly Ser Leu Thr Ser Thr Ser 410 Glu Gln Ala Gln Leu Ala Thr Ser Glu Ser Leu Ser Pro Leu Thr Ser 420 425 430 Leu Ser Gly Gln Ala Pro Leu Ala Thr Gly Glu Ser Leu Ser Asp Leu 440 Pro Ser Thr Ser Glu Gln Ala Arg His Ser Tyr Gly Val Arg Phe Asn 450 Gly Phe Gln Ala Asp Asp Ser Ile Phe Pro Thr Glu Ile Arg Asn Arg Val Glu Ala His Gly His Gly Val Thr His Asp His Glu Asp Ser Asn 485 490 Glu Ser Leu Ser Ser Asp Glu Arg His Gly His Gly Pro Ser Gly Lys 500 505 Pro Met Leu His His Gly Glu Lys Gly Val Gln Glu Ala Gly Trp Asp 520 Leu Asp Asp Asn Asn Asp Lys Ser Asp Cys Leu Ala Ile Lys Glu Gln 530 535 Phe Lys Cys Asp Thr Asn Ser Thr Trp Gly Leu Asn Asp Asp Glu Leu Met Ala His Gly Gln Glu Lys Asp Ser Ser Ser Glu Ser Glu Asp Ser

570

- 34 -Cys Pro Pro Ser Pro Gly Cys Ser Phe Thr Glu Gly Phe Ser Phe Asp 585 Leu Phe Asn Pro Asp Tyr Val Pro Lys Val Asp Lys Trp Ser Arg Phe 600 Leu Phe Pro Leu Ala Phe Gly Leu Phe Asn Ile Val Tyr Trp Val Tyr 610 615 His Met Tyr 625 (2) INFORMATION FOR SEQ ID NO: 5: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 19 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: cDNA (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5: GCAAATGAAG CTGTGGTTC 19 (2) INFORMATION FOR SEQ ID NO: 6: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 20 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: cDNA (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6: CAATGTTGAA CAACCCAAAG 20 (2) INFORMATION FOR SEQ ID NO: 7:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 19 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7:	
GCCTGAGACC GAATTTTGG	19
(2) INFORMATION FOR SEQ ID NO: 8:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 19 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 8:	
GGAACCGGGA CCACTTGTC	19
(2) INFORMATION FOR SEQ ID NO: 9:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 18 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 9:	
TAGTCCAGGG TCAAGTTC	18
(2) INFORMATION FOR SEQ ID NO: 10:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 20 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	·
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 10:	
TAGTATGCTA AGCGTGAATC	20

(2) INFORMATION FOR SEQ ID NO: 11:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 18 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 11:	
GAGTTTGAGG ATAGTTGC 18	
(2) INFORMATION FOR SEQ ID NO: 12:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 18 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single	
(D) TOPOLOGY: linear (ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 12:	
TGCTCCTTCA CTGAAGGG 18	
(2) INFORMATION FOR SEQ ID NO: 13:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 42 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 13:	
CCATGACTCA AGCTTGCCAC CATGCTGCGA GCCGCAGTGA TC	42
(2) INFORMATION FOR SEC ID NO. 14.	

(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 27 base pairs

- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 14:

TGAAAGGAGC ACAGCACAGT GCTCCCG

27

- (2) INFORMATION FOR SEQ ID NO: 15:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 45 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: cDNA
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 15:

CTGCTTCTTG CACACCCTTC TCGCCATGGT GAAGCATGGG CTTCC 45

- (2) INFORMATION FOR SEQ ID NO: 16:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 45 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: cDNA
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 16:

TGTCGCCTAG GCTGGCGCCG AGGTCCTCGA CTGTAGAAAA GATAG 45

CLAIMS:

- 1. A stably co-transfected eukaryotic cell line capable of expressing a GABA receptor, which receptor comprises the theta receptor subunit, at least one alpha receptor subunit and optionally one or more receptor subunits selected from the beta, gamma, delta and epsilon subunits.
- 2. A cell line according to claim 1 which is a rodent fibroblast 10 cell line.
 - 3. A process for the preparation of an eukaryotic cell line capable of expressing a GABA receptor, which comprises stably cotransfecting a eukaryotic host cell with at least two expression vectors, one such vector harbouring the cDNA sequence encoding the theta GABA receptor subunit, another such vector harbouring the cDNA sequence encoding an alpha GABA receptor subunit, and optionally one or more additional vectors harbouring the cDNA sequence encoding a beta, gamma, delta or epsilon GABA receptor subunit.

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- 4. A process according to claim 3 wherein the cell line is a rodent fibroblast cell line.
- 5. A DNA molecule encoding the theta subunit of the human

 GABA receptor comprising all or a portion of the sequence depicted in

 Figure 1 or Figure 2, or a modified human sequence.
- 6. A recombinant expression vector comprising the nucleotide sequence of the human GABA receptor theta subunit together with additional sequences capable of directing the synthesis of the said human

WO 98/49293 - 39 - PCT/GB98/01206

GABA receptor theta subunit in cultures of stably co-transfected eukaryotic cells.

- 7. A protein preparation of GABA receptor subunit combinations derived from a cell line according to claim 1 or 2.
 - 8. A membrane preparation containing subunit combinations of the GABA receptor derived from a cell line according to claim 1 or 2.
- 9. A preparation according to claim 7 or 8 wherein the subunit combination derived is the $\alpha_1\theta\gamma_2$, $\alpha_2\beta_1\theta\gamma_1$ or $\alpha_2\beta_3\theta\gamma_2$ subunit combination of the GABA receptor.
- The use of a cell according to claim 1 or 2 or a membrane
 preparation derived therefrom in screening for and designing medicaments
 which act upon a GABA receptor comprising the theta subunit.
 - 11. A method for determining whether a ligand, not known to be capable of binding to a human GABAA receptor comprising the theta subunit, can bind to a human GABAA receptor comprising the theta subunit, which comprises contacting a mammalian cell comprising DNA molecules encoding at least one alpha receptor subunit, the theta receptor subunit and optionally one or more beta, gamma, delta or epsilon receptor subunits, with the ligand under conditions permitting binding of ligands known to bind to the GABAA receptor, detecting the presence of any of the ligand bound to the GABAA receptor comprising the theta subunit and thereby determining whether the ligand binds to the GABAA receptor comprising the theta subunit.
- 30 12. A method of screening drugs to identify drugs which specifically interact with, and bind to, a human GABAA receptor

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comprising the theta subunit on the surface of a cell which comprises contacting a mammalian cell comprising a DNA molecule encoding at least one alpha receptor subunit, the theta receptor subunit and optionally one or more beta, gamma, delta or epsilon receptor subunits, on the surface of a cell with a plurality of drugs, determining those drugs which bind to the mammalian cell, and thereby identifying drugs which specifically interact with, and bind to, human GABAA receptors comprising the theta subunit.

- 13. A polynucleotide which hybridizes under stringent conditions10 to the DNA molecule depicted in Figure 1 or Figure 2.
 - 14. A GABA_A receptor theta subunit polypeptide which has the deduced amino acid sequence of Figure 1 or Figure 2, or a fragment, analog or derivative thereof.

FIGURE 1

<u>Human θ Subunit</u>

Τ.	ATGCTGCGAGCCGCAGTGATCCTGCTCATCAGGACCTGGCTCGCC	GGAGG	GCAACTAC	. 60
	M L R A A V I L L I R T W L A	E G	N Y	
61	CCCAGTCCCATCCCGAAATTCCACTTCGAGTTCTCCTCTGCTGTGCCC	רכא א כי	maamaama	
	PSPIPKFHFEFSSAVP	E V	V L	120
121	AACCTCTTCAACTGCAAAAATTGTGCAAATGAAGCTGTGGTTCAAAAC	ייייייי עיבי	TCC3 C3 CC	100
	N L F N C K N C A N E A V V Q K	I L	D R	180
181	GTGCTGTCAAGATACGATGTCCGCCTGAGACCGAATTTTGGAGGTGCC	accerci	TOOOMOMO	
	V L S R Y D V R L R P N F G G A	-CC1G	P V	240
243				
241	AGAATATCTATTTATGTCACGAGCATTGAACAGATCTCAGAAATGAAT	[ATGG]	ACTACACG	300
	RISIYVTSIEQISEMN	M D	YT	
301	ATCACGATGTTTTTCATCAGACTTGGAAAGATTCACGCTTAGCATAC	TATG	AGACCACC	360
	ITMFFHQTWKDSRLAY	Y E	ТТ	300
361				
201	CTGAACTTGACCCTGGACTATCGGATGCATGAGAAGTTGTGGGTCCCT	GACTO	CTACTTT	420
	LNLTLDYRMHEKLWVP	D C	Y F	
421	CTGAACAGCAAGGATGCTTTCGTGCATGATGTGACTGTGGAGAATCGC	GTGT1	TCAGCTT	480
•. •	LNSKDAFVHDVTVENR	V F	ОГ	
481	CACCCAGATGGAACGGTGCGGTACGGCATCCGACTCACCACTACAGCA	ርጥጥጥር	ייייירריייר	540
	HPDGTVRYGIRLTTA	V C	S T.	540
541	GATCTGCATAAATTCCCTATGGACAAGCAGGCCTGCAACCTGGTGGTA	GAGAG	CTATGGT	600
	DLHKFPMDKQACNLVV	E S	Y G	
601	TACACGGTTGAAGACATCATATTATTCTGGGATGACAATGGGAACGCC	מייירמ	<u>ር</u> ውጥር እ ርጥ	660
	YTVEDILFWDDNGNA	I H	M T	860
661	GAGGAGCTGCATATCCCTCAGTTCACTTTCCTGGGAAGGACGATTACT	AGCAA	GGAGGTG	720
	EELHIPQFTFLGRTIT	s ĸ	E V	
721	TATTTCTACACAGGTTCCTACATACGCCTGATACTGAAGTTCCAGGTTC	CAGAG	GGAAGTT	780
•	YFYTGSYIRLILKFQV	Q R	E V	780
781	AACAGCTACCTTGTGCAAGTCTACTGGCCTACTGTCCTCACCACTATTA			
	N S Y L V Q V Y W P T V L T T I T	ACCTC	TIGGATA	840
341		TTAAC'	TTCAATG	900
	SFWMNYDSSAARVTIGI	L T	s M	
901	CTCATCCTGACCACCATCGACTCACATCTGCGGGATAAGCTCCCCAACA	ል ጥጥጥ ሌ.	ርጥር ጥ አ ጥር	0.00
	LILTTIDSHLRDKLPNI	I S	C I	960
961			•	
, , , ,	AAGGCCATTGATATCTATATCCTCGTGTGCTTGTTTCTTGTGTTCCTGT K A I D I Y I L V C L F F V F L S	CCTT	GCTGGAG	1020
		s I.	1. F	

1021	T	ATG'	TCT	ACA	TCA	ACT	ATC	TTT	TĊT	ACA	GTC	GAG	GAC	СТСС	GGC(GCC	AGC	רידא	acc	GACGC	3.000
	Y	V	Y	I	N	Y	L	F	Y	s	R	G	P	R	R	Q	P	R	R	R	1080
1081	A	GGA(GAC	CCC	GAA	GAG	TCA'	TTG	CCC	GCT.	ACC	GCT2	ACC	AGC	AAG'	TGG'	TGG	ТАС	ממבו	ACGTG	1140
	R	R	P	R	R	V	I	A	R	Y	R	Y	Q	Q	V	v	v	G	N	V	1140
1141	C	AGG	ATG	GCC'	TGA	TTA	ACG'	TGG	AAG.	ACG	GAG:	CAC	CTO	TCI	rcco	CA'	רכם	ריי. ריירי	: CDG(CCCAC	1200
	Q	D	G	L	Ι	N	V	E	D	G	v	S	s	L	P	I	T	P	A	Q	1200
1201	G	CCC	CCC	rgg	CAA	GCC	CGG	AAA	GCC'	rcg	GTTC	CTTT	GAC	GTC	CAC	CTC	CG	AGC	AGGG	CCCAG	1260
	A	P	L	A	s	P	E	s	L	G	s	L	T	s	T	s	E	Q	A	Q	1200
1261	C	rggc	CAC	CTC	CGG	AAA(GCC:	CAC	3CC	CAC	rcac	TTC	TCT	CTC	AGO	GCZ	AGG	CCC	CCI	rggcc	1320
	L	A	Т	S	E	S	L	S	P	L	T	S	ŕ	S	G	Q	A	P	L	A.	2320
1321	ΑC	TGG	AGA	AAC	GCC.	rga(3CG <i>I</i>	ATCI	יככי	CTC	CAC	CTC	'AGA	GCA	GGC	:ccc	GC.	ACAC	СТА	ላፐርርጥ	1380
	T	G	E	S	L	s	D	L	P	s	Т	s	E	Q	A	R	Н	s	Y	G	1300
1381	GT	TCG	CTI	'TA	ATGO	TTT	rcc#	AGGC	TG	ATGA	CAG	TAT	TAT	TCC	TAC	CGA	AAI	race	CAA	CCGT	1440
	V	R	F	N	G	F	Q	A	D	D	s	I	I	P	T	E	I	R	N	. R	1110
1441	Gī	'CGA	AGC	CCA	TGG	CCA	TGG	TGI	TAC	CCA	TGA	.CCA	TGA	AGA	TTC	CAA	TGA	GAG	CTT	GAGC	1500
•	V	E	A	H	G	Н	G	V	T	H	D	Н	E	D	s	N	E	s	L	s	1300
1501	TC	GGA	TGA	.GCG	CCA	TGG	IĆCA	TGG	CCC	CAG	TGG	GAA	GCC	CAT	GCT	TCA	.CCA	TGG	CGA	GAAG	1560
	S	D ·	E	R	H	G	H	G .	P	S	G.	K	P	M	L	H	H	G	E	K	
1561	GG	TĠT	GCA	AGA	AGC	AGG	CTG	GGA	CCI	'TGA	TGA	CAA	CAA	TGA	CAA	GAG	CGA	CTG	CCT	TGCC	1620
	G	V	Q	E	A	G	W	D	L	D	D	N	N	D	K	S	D	С	L	A	
1621	AT	TAA	GGA	GCA	ATT	CAA	GTG	TGA	TAC	TAA	.CAG	TAC	CTG	GGG	CCT	TAA	TGA	TGA	TGA	GCTC	1680
	Ι	K	E	Q	F	K	С	D	Т	N	S	Т	W	G	L	N	D	D	E	L	
1681	GT	GGC	CCA'	TGG	CCA	AGA	GAA	GGA	CAG	TAG	CTC	AGA	GTC:	rga(GGA'	TAG'	TTG	CCC	CCC	AAGC	1740
	V	A	H	G	Q	E	K	D	S	S	S	Ē	s	E	D				P		2710
1741	CC	IGG	GTG	CTC	CTT	CAC	TGA	AGG	GTT	CTC	CTT	CGA:	rcto	CTT	raa'	rcc'	TGA	СТА	ССТ	CCCA	1800
	P	G	С	s	F	T	E	G	F	s	F	D	L	F	N	P	D	Y	V	P	1000
1801	AA	GGT	CGA	CAA	GTG	GTC	CCG	GTT	ССТ	CTT	CCC:	rcto	3GC(TTT	rgg	STT	3TT	CAA	CAT'	TGTT	1860
	K	V	D	K	W	s	R	F	Ļ	F	P	L	A	F	G	L,	F	N	r		
1861	TA	CTG	GT	ATA	CCA'	TAT	GTA'	TTA	3		1884	1									

FIGURE 2

Human θ Subunit

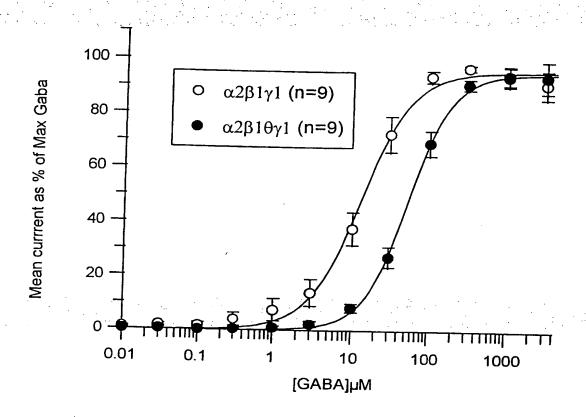
1	7	ATG	CTG	CGA	GCC	GCA	GTG.	ATC	CTC	CTC	CT	CA:	TCA	GGA	CC	TGG	CTC	GÇG	GA	GGG	CA	ACTA	C 6
		4	L	R ·	A .	A 1	V	1	L:	L	L	I	R	τ	' '	W	L ·	A	E	G	N	Y	
61	c	ccc	AGT	CCC.	ATC	CCG	\AA?	rtc	CAC	TTC	GA	GTI	CT	ССТ	CTO	3CT(GTG	CCC	GAZ	AGT	'CG'	rccto	G 120
						PF									7							L	
121	A	ACC	CTC'	TTC	\AC7	rgca	\AAA	\AT7	rg T	GCA	AAT	ГGA	AG	CTG	TGG	TTC	CAA	AAG.	АТТ	TT	GG <i>I</i>	CAGO	; 180
						СК																R	
181	G	TGC	TG	CAA	GAI	ACG	ATG	TCC	:GC	CTG.	AGA	ACC	GAZ	TT:	ГТG	GAG	GTO	CCC	CT	GT	GCC	TGTG	240
						D															P	v	
241	A	GAA	TAT.	CTA	TTT	ATG	TCA	CGA	.GCI	ATT	GAA	CA	GAI	CTO	CAG	ААА	TGA	ATA	\TG	GA	СТА	CACG	300
	R	Ι	٤	I	Y	v	Т	s	1	[]	3	Q	ı	s	E	M	N	1 1	1	D	Y	Т	
301																						CACC	360
÷	1	T	. M	F	F	Н	Q	T	W		C :	D	S	R	L	A	Y	Y				T	. ,
361						rgg <i>i</i> D																CTTT	-420
																						F	
121																						CTT	480
	_	••	5		ם	A	r	V	н	D	' 1	,	Т	V	Е	N	R	V	F	•	Q	L	
81																TAC	AG	CAG	CTT	GT	TCC	CTG	540
	Н	P	D	G	T	V.	R	Y	G	I	F	3	L	T	Т	T	A	A	С		S	L	
41																				GC.	ГАТ	GGT	600
	D	L	н	ĸ	F	P	М	D	K	Q	A	. (С	N	L	. V	v	E	s	3	Y	G	
01						CAT		ATT	ATI	CT	GG	ATO	GAC	'AA'	rgg	GAA	CGC	CA1	CC.	AC!	\TG	ACT	660
	Y	Т	V	E	D	Ι	1	L	F	W	D	• 1	D ·	N	G	N	A	I	H	1	1	T	
61	GAG	GGA	GCT	GCA	TAT	CCC:	rca(GTT	CAC	TT1	CC	TGO	G A	AGG	AC	GAT	TAC	TAG	CA	AGG	ag	GTG	720
	E	E	L	Н	I	P	Q	F	T	F	L	C	3	R	T	I	T	s	ĸ	F	,	v	
21	TAT	TT	CTA	CAC	AGG'	rtco	TAC	CAT	ACG	CCI	'GA'	TAC	TG.	AAG	TT	CA	GGT	TCA	GAC	GGG	AA	3TT	780
	Y	F	Y	T	G	S	Y	I	R	L	I	I	<u>.</u> :	ĸ	F	Q	v	Q	R	E	: 1	V	
31	AAC	AGO	TA	CCT:	rgto	CAA	GTC	TAC	TG	GCC	TAC	CTG	TC	CTC	ACC	CAC	TAT	TAC	CTO	TT	GG2	ATA	840
	N	s	Y	L	v	Q	v	Y	w	Þ	т	1,7	, ,	٠	т	T	т	m	c	7.7			

841	T	'CG	rtt:	TGG#	ATG	AC:	CATO	SAT	rcci	CTC	GCAC	3CC2	AGG	GTG.	ACA	ATT	GGC	ттл	ACI	rrc	CAATG	900
	S			W I			ľ				A 1		۲ ۲			I (300
901	C	TC	ATCO	TGF	ACC	CCZ	TCG	ACI	CAC	ATC	TGC	GGG	SATA	LA G(CTC	ccci	AĄC:	ATT	TCC	TC:	STATC	960
	L		I	T .	ר ז	' I			H	I	F	ł . E	ŀ	(1	ւ	Pì	1 :	T .	S	С	r ,	
961	A	AGG	CCA	TTG	АТА	TCI	'ATA	TCC	TCC	TCT	יככז	יייביי	` `	ייייי	·	rr.c.c	-		nma	-	GGAG	
																; I						1020
1021	T	ATG	TCT	'ACA	TCA	ACT	ATC	TTT	TCT	ACA	GTC	GAG	GAC	CTC	GGG	GCC	AGC	CT	AGG	CG	ACAC	1080
																R Q						
1081	AC	GA	GAC	CCC	GAA	GAG	TCA	TTG	CCC	GCT	ACC	GCT.	ACC.	AGC	AAG	TGG	TGG	TAC	GA	AA.	CGTG	1140
	R	R	P	R	R	v	I	A	R	Y	R	Y	Q	Q	V	v	ν		3 1	N	v	
1141	CA	\GG	ATG	GCC.	rga:	rta.	ACG:	rgg	AAG	ACG(GAG:	rca(GCT	CTC	TCC	CCA	TCA	ccc	CAC	GC	GCAG	1200
	Q	D	G	L	I	N	v	E	D	G	v	s	S	L	P	I	T	P	, ,	4	Q	
1201									GCC1	rcgo	STTC	TT	rga(CGT	CCA	CCT	CCG.	AGC	AGC	cc	CCAG	1260
	A	Р	L	A	s	P	E	s	L	G	s	L	Т	S	Т	s	E	Q	P	4	Q	
1261	CT	GG	CCAC	СТС	GG#	AAC	CCI	CAC	CCC	ACI	CAC	TTC	TC	CT	CAG	GCC	AGG	ECC	cçc	TC	GCC	1320
	L	A	T	S	E	s	L	s	P	L	T	s	Ĺ	s	G	Q	A	P	L	• [.	A	
321	AC	TGG	SAGA	LAA G	CCI	'GAC	CGA	TCI	ccc	CTC	CAC	CTC	'AGA	.GCZ	4GG(ccc	GC2	ACA(GCT	'A'	GGT	1380
	T	G	E	s	L	s	D	L	P	s	T	s	E	Q	A	R	н	s	Y		G	
.381	GT"	TCG	CTT	TAA	TGG	ттт	CCA	.GGC	TGA	TGA	.CAG	TAT	TTI	TCC	TAC	CCGA	LAAT	rcc	GCA	AC	CGT	1440
	V	R	F	N	G,	F	Q	A	D	D	s	I	F	P	T	E	I	R	N		R	
441	GT	CGA	AGC	CCA	TGG	CCA	TGG	TGT	TAC	CCA	TGA	CCA	TGA	AGA	TTC	CAA	TGA	GAG	CT	TG	AGC	1500
	v	E	A	H	G	H	G	v	T	н	D	Н	E	D	s	N	E	s	L		s	
501	TCC	GA.	TGA	GCG	CCA	TGG	CCA	TGG	ccc	CAG'	TGG	GAA	GCC	CAT	GCI	TCA	.CCA	TGO	3CG	AG.	AAG	1560
•																Ħ						
561	GGT	GT	GCA	AGA	AGC	AGG	CTG	GGA	CCT	rga:	TGA	CAA	CAA'	TGA	ממי	.GAG	CCY	CTO		r-r	GCC	1620
																s						1620
621	ATI	'AAC	GGA	GCA	ATT	CAAC	GTG:	rga:	rac:	raac	CAG	rac	CTG	GGG	CCT	TAA	TGA	TGA	TG	A.G.	CTC	1680
																N						1000
681	ATG	GC	CCA?	rggo	CA	\GA(SAAC	GGAC	CAG	rag(CTC	\GA(STC:	rga(GGA	TAG'	rtg:	ccc	ccc	CAJ	AGC	1740
												_	_	_	_	_	_	_				

- 1741 CCTGGGTGCTCCTTCACTGAAGGGTTCTCCTTCGATCTCTTTAATCCTGACTACGTCCCA 1800
 P G C S F T E G F S F D L F N P D Y V P
- 1801 AAGGTCGACAAGTGGTCCCGGTTCCTCTCCCTCTGGCCTTTGGGTTGTTCAACATTGTT 1860

 K V D K W S R F L F P L A F G L F N I V
- 1861 TACTGGGTATACCATATGTATTAG 1884

FIGURE 3.



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PCT/GB 98/01/206

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Fax: (+31-70) 340-3016 Le Cornec. N

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